

Antibacterial Activity and Structure Elucidation of Salicin from Stem Bark of *Salix tetrasperma* ROXB.

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Abstract

Salix tetrasperma Roxb. (Family Salicaceae) is a plant that used as traditional medicine for anti-inflammatory, analgesic, reduces fever, and itching medicine. In this study was carried out extraction, isolation, structure elucidation of salicin from *Salix tetrasperma* Roxb. stem bark and it's antibacterial activity. The extraction method was used the maceration method by *n*-hexane, ethyl acetate, and methanol solvents. Isolation of compound from ethyl acetate extract of *Salix tetrasperma* Roxb. stem bark using chromatography methods and obtained white solid (15 mg). The structure was elucidated using spectroscopic analysis, including *Ultraviolet* (UV), *Infrared* (IR), *Nuclear Magnetic Resonance* (NMR) and comparative literature, identified as salicin compound with molecule formula C₁₃H₁₈O₇. Antibacterial activity was evaluated against *Escherichia coli* and *Staphylococcus aureus* bacteria using disk diffusion method. This compound has a great an antibacterial activity against *Staphylococcus aureus* bacteria with clear zone diameter of 10.2 ± 0.3 mm. This shows that the *Salix tetrasperma* Roxb. stem bark has great potential as a source of antibacterial compound.

Keywords: *Salix tetrasperma* Roxb., salicin, antibacterial activity

Abstrak (Indonesian)

Salix tetrasperma Roxb. (famili salicaceae) adalah salah satu tumbuhan yang digunakan sebagai obat anti-inflamasi, analgesik, mengurangi demam dan obat gatal-gatal. Pada penelitian ini dilakukan ekstraksi, isolasi, elusidasi struktur senyawa salisin dari kulit batang *Salix tetrasperma* Roxb. dan aktivitas antibakterinya. Metode ekstraksi yang digunakan adalah maserasi menggunakan pelarut *n*-heksana, etil asetat dan metanol. Isolasi senyawa menggunakan metode kromatografi diperoleh padatan putih (15 mg). Elusidasi struktur menggunakan analisa Spektroskopi *Ultraviolet* (UV), *Infrared* (IR), *Nuclear Magnetic Resonance* (NMR) dan literatur pembandingan, diidentifikasi sebagai senyawa salicin dengan rumus molekul C₁₃H₁₈O₇. Uji aktivitas terhadap bakteri *Escherichia coli* dan *Staphylococcus aureus* menggunakan metoda *disk diffusion* menunjukkan aktivitas antibakteri yang baik terhadap bakteri *Staphylococcus aureus* dengan diameter zona bening sebesar 10.2 ± 0.3 mm. Ini menunjukkan bahwa kulit batang *Salix tetrasperma* Roxb. memiliki potensi yang baik sebagai sumber senyawa antibakteri.

Kata Kunci: *Salix tetrasperma* Roxb., salicin, aktivitas antibakteri

INTRODUCTION

Salix tetrasperma Roxb. plant (Family; Salicaceae) commonly called Indian Willow [1].

In Indonesia, it's known as dalu- dalu used as traditional medicine. The phenolic glycosides contained in *Salix tetrasperma* Roxb. reported as

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an anti-inflammatory, analgesic, reduce fever and reduce rheumatic infection, headache, the chronic pain syndrome[2], cough, scorpion sting, bug bite, wounds, warts[3], and dysmenorrhea in women[1].

Various bioactivities have been reported of *Salix tetrasperma* Roxb. plant including anti-inflammatory, analgesic [4], antioxidant [4,5], antiprotozoa [6], hypoglycemic [7], diuretic, laxative [8], cytotoxic [9,10], antischistosoma [11], insecticidal [12], antifungal [13], and antibacterial activities [9].

Phytochemical investigation from the bark extract of this plant has been reported steroid, sterol, triterpene, tannin, phenolic compounds, saponin, and flavonoid [7, 8]. Research conducted at Zagazig University, Egypt has been reported that the *Salix tetrasperma* Roxb. plant generated some pure compound, including β -sitosterol acetate, friedelin, 3β -friedelinol, β -amyrin, β -sitosterol, β -sitosterol-*O*-glucoside, and palmitic acid, which has been isolated from the methanol extract of *Salix tetrasperma* Roxb. from dichloromethane fraction of this leaf has been isolated catechol and tremulacin. Salicin and its derivatives tremuloidin and 2'-*O*-*p*-(*E*)-coumaroyl salicin were isolated from the ethyl acetate fraction of the leaf [4].

This paper reports the salicin isolated compound from ethyl acetate extract of *Salix tetrasperma* Roxb. stem bark. Structure of the salicin compound was elucidated using spectroscopic analysis, including ultraviolet visible spectrophotometer, FT-IR spectrophotometer and Nuclear Magnetic Resonance spectrometer. Further, Antibacterial activity from the salicin compound, *n*-hexane, ethyl acetate, and methanol extracts of *Salix tetrasperma* Roxb. stem bark was evaluated using disk diffusion method against *Escherichia coli* and *Staphylococcus aureus* bacteria.

MATERIALS AND METHODS

Materials

Plant material

The stem bark of *Salix tetrasperma* Roxb. was collected from Pesisir Selatan Regency, West Sumatera, Indonesia. The sample has been identified in the Herbarium of Andalas

University (ANDA), Department of Biology, Faculty of Mathematics and Natural Sciences, Andalas University with the specimen code 237/K-ID/ANDA/V/2018 and collection number SR-01.

Media and Chemical material

The media and chemicals used in the research were Mueller-Hinton agar (Hi-media), dimethyl Sulfoxide (DMSO), NaCl 0.9%, distilled methanol, acetone, ethyl acetate, dichloromethane, *n*-hexane (Brataco) as the solvents, H₂SO₄ 2N, silica gel 60 (Merck, 0.063-0.200 mm), TLC plate (Merck, DC-Alufolien Kiesegel 60 F₂₅).

Instrument

Distillation apparatus, macerator, Rotary Evaporator (Heidolph VV 2000), melting point apparatus (Fisher-Johns), oven, vacuum desiccator, scales, UV lamps GL-58 (λ 254 and 365 nm), ultraviolet visible spectrophotometer (Thermo Scientific, Genesys 10 s UV-Vis), FT-IR spectrophotometer (Perkin Elmer, Frontier), Nuclear Magnetic Resonance (NMR) spectrometer (JEOL JNM-ECZ500R), column chromatography, petri dish, disc paper, laminar flow and commonly used glassware in laboratories.

Methods

Extraction

The stem bark of *Salix tetrasperma* Roxb. was dried in the shade and finally ground to a powder. The dry powder stem bark of *Salix tetrasperma* Roxb (6 kg) was extracted by maceration method using *n*-hexane, ethyl acetate, and methanol solvents at room temperature. It was obtained crude extracts of *n*-hexane (20 g), ethyl acetate (33 g) and methanol (250 g).

Isolation and Purification

The ethyl acetate extract (20 g) was applied on to silica gel of vacuum liquid chromatography, packed with *n*-hexane. The polarity of the eluent was increased gradually using *n*-hexane-ethyl acetate (8:0 to 2:10) and ethyl acetate:methanol (10:0 to 0:10). Each fraction in TLC and fractions the same spot

pattern were combined to obtain 10 fractions (A-J). Further, the fraction E (2.5 g) was separated by using gravity column chromatography technique with a gradient elution system using silica gel as stationary phase and *n*-hexane:acetone (7:0 to 3:10) as a mobile phase. The same pattern of every spot on TLC was combined to obtain 10 sub-fractions (E₁- E₁₀). E₅ was purified with acetone obtained 15 mg white solid. Further, the melting point of the isolated compound was measured and characterization of the isolated compound using UV, IR, NMR spectroscopy.

Test bacteria and Inoculum Preparation

The *Escherichia coli* and *Staphylococcus aureus* bacteria were cultured in tilted Nutrient Agar media for 24 hours and incubated at 37 °C, then taken loopful and suspended in a tube containing 200 µL of physiologic salt solution (NaCl 0.9%). Sterile liquid Mueller-Hinton agar was added to 20 ml of each petri dish and allowed to solidify at room temperature. The media was dripped with 200 µL of the bacterial suspension tested and flattened with an L stem, then left to dry for 15 minutes in a laminar flow [14].

Antibacterial activity

Sterile disc paper with a diameter of 6 mm added *n*-hexane, ethyl acetate, methanol solvents and salicin compound (20 µL). The concentrations of each extract are 1000, 500 and 250 µg / mL. Further, the paper discs placed on MHA media and incubated at room temperature for 24 hours. Positive control used Amoxicillin 250 µg / mL and negative control used DMSO 100% and methanol. The clear zone around the disc showed the area of bacterial resistance. The diameter of the clear zone measured horizontally and vertically using a scale ruler [15].

RESULTS AND DISCUSSION

Isolation and Purification

Phytochemical profile study of ethyl acetate extract of *Salix tetrasperma* Roxb. showed that

the stem bark of this plant containing phenolic group, flavonoid, terpenoid, steroid, and saponin.

Phytochemical profile study of ethyl acetate extract of *Salix tetrasperma* Roxb. showed that the stem bark of this plant containing phenolic group, flavonoid, terpenoid, steroid, and saponin. Purification using chromatography method obtained 15 mg of white solid. The TLC result showed single pink spot after added H₂SO₄ 2N. The melting point of the isolated compound was 189-190 °C. The ultraviolet spectrum in methanol showed absorption at 223 nm and 271 nm with the presence of a phenolic moiety. Infrared spectrum showed absorption at (cm⁻¹) 3317.09 (OH); 2921.39 (CH); 1593.52 (C=C aromatic); and 1021.65 (C-O) cm⁻¹.

¹H-NMR (CD₃OD), 500 MHz (ppm) spectra data salicin compound showed the value of chemical shift at 7.34 (1H); 7.26 (1H); 7.22 (1H); 7.03 (1H); 4.87 (1H); 4.79 (1H); 4.56 (1H); 3.91 (1H); 3.71 (1H); 3.51 (1H); 3.46 (1H); 3.42 (1H) and 3.39 (1H).

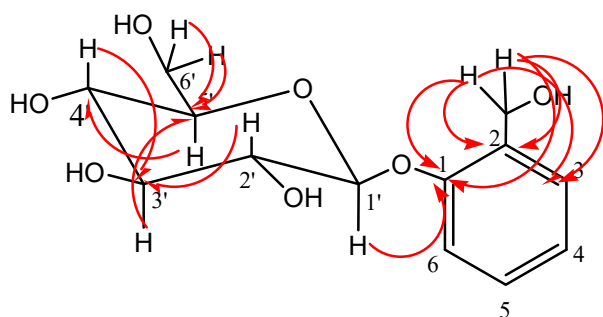
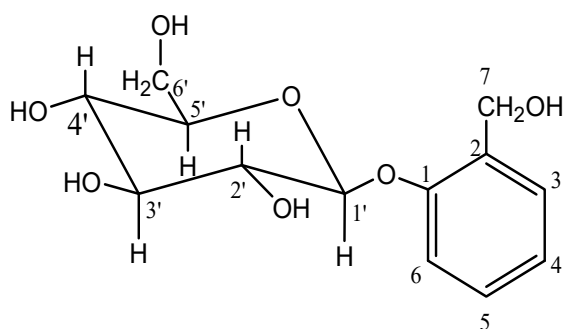
¹³C-NMR (CD₃OD), 125 MHz (ppm); spectra data Salicin compound showed the value of chemical shift at 61.0; 62.6; 71.4; 75.1; 78.0; 78.3; 103.4; 117.1; 123.7; 129.9; 130.0; 132.2 (ppm). ¹H-NMR and ¹³C-NMR of isolated compound result showed thirteen minimum C atoms supported by HSQC spectrum data. Which appear as two secondary C atoms, nine tertiary C atoms and two quaternary C atoms. This spectrum showed the relationship between ¹H-¹³C. Proton H-3, H-4, H-5 and H-6 were aromatic ring protons bound to C-3, C-4, C-5 and C-6 atoms which were located in the chemical shift C=C.

HBMC correlation, shown in Figure 1, correlation between H-7 and H-7b with atoms C-1, C-2, and C-3; H-1' with atom C-1; H-2' with atom C-3'; H-3' with atom C-5'; H-4' with atom C-3'; H-5' with atom C-4'; and H-6' with atom C-5'. This proved there was a correlation between protons and carbons.

Table 1. ^1H (CD_3OD ; 500 MHz), ^{13}C (CD_3OD ; 125 MHz) NMR data of the isolated compound and NMR data comparative of salicin compound in $\text{D}_2\text{O}:\text{CD}_3\text{OD}$ (4:1) by Dias [16].

No	δ_{C} (ppm)	δ_{H} (ppm)	$J_{\text{H-H}}$ (Hz); multiplicity	HMBC	δ_{C} (ppm) comparative	δ_{H} (ppm) comparative
1	157.2	-	-		157.6	
2	132.2	-	-		133.9	
3	130.0	7.26	8.2; 1.4; (td)	C ₁ ; C ₅	132.1	7.5 (dd)
4	123.7	7.03	8.2; 0.9; (td)	C ₂ ; C ₆	126.0	7.15 (td)
5	129.9	7.34	7.6 (d)	C ₁ ; C ₃	132.3	7.37 (td)
6	117.1	7.22	7.9 (d)	C ₁ ; C ₂ ; C ₄	118.0	7.21 (d)
7	61.0	4.56(1H) 4.79(1H)	12.9 (d) 12.9 (d)	C ₁ ; C ₂ ; C ₃ C ₁ ; C ₂ ; C ₃	62.0	4.68 (d) 4.73 (d)
1'	103.4	4.87	7.8 (d)	C ₁	103.4	5.08 (d)
2'	75.1	3.42	overlapped	C _{3'}	75.7	3.56- 3.63
3'	78.0	3.51	overlapped	C _{5'}	78.6	3.56- 3.63
4'	71.4	3.39	(m)	C _{3'}	72.1	3.5 (m)
5'	78.3	3.46	(m)	C _{4'}	78.6	3.56-3.63 (m)
6'	62.6	3.71 (1H) 3.91 (1H)	11.9; 5.3; (dd) 11.9; 1.8 (dd)	C _{5'} C _{5'}	63.3	3.75 (dd) 3.91 (dd)

Based on ^{13}C -NMR, ^1H -NMR, HSQC, HMBC spectroscopy data and chemical shift data of isolated of compounds with the salicin compound reported data by Dias showed high suitability, Table 1 [16]. It can be concluded that isolated compound was salicin with molecular formula $\text{C}_{13}\text{H}_{18}\text{O}_7$ as shown in Figure 2.

**Figure 1.** ^1H - ^{13}C HMBC correlation of Salicin**Figure 2.** Structure of Salicin

Antibacterial Activity

Antibacterial activity of *Salix tetrasperma* Roxb is listed in Table 2. The Table showed the ethyl acetate extract has the largest clear zone diameter compared with the other extracts for *Staphylococcus aureus* bacteria (14.5 ± 0.5 mm), followed by methanol extract (12.7 ± 0.6) and salicin compound (10.2 ± 0.3) for *Escherichia coli* bacteria, the largest clear zone diameter was found in *n*-hexane extract (11.7 ± 0.8 mm).

Difference of active extract against each bacteria was caused by differences in secondary metabolites that dominant content in each extract. The ethyl acetate extracts are dominant contains phenolic and flavonoid compounds. Phenol and polyphenol compounds are largest groups of secondary metabolites, has hydroxyl groups attached at aromatic phenol group [17]. Phenol can be to change the permeability of bacterial cells, caused them lost the macromolecule of the cells. The compound also disrupts membrane function and affect membrane protein caused that change in structure and function. At low concentration, phenolic compound affects enzyme activity and at high concentration, causes protein denaturation [18]. While for *Escherichia coli* bacteria, the extract that has higher inhibitory was *n*-hexane extract. In *n*-hexane extract,

secondary metabolites the most dominant of a compound terpenoid. Terpenoid was an organic compound that disrupts the formation of membranes by the lipophilic compound. The

ability of terpenoid damage the cell membrane, deactivate enzyme and protein denaturation causes that permeability of bacterial cell walls to decrease, and cell walls are damage [19].

Table 2. Diameter of clear zone bark extract of *Salix tetrasperma* Roxb. and salicin compound against bacterial growth of *Escherichia coli* and *Staphylococcus aureus* bacteria

Extract	Concentrations (ppm)	Diameter of suppression zone (mm)	
		<i>Escherichiacoli</i>	<i>Staphylococcus aureus</i>
Methanol	250	7.8 ± 0.3	9.2 ± 0.3
	500	8.3 ± 0.3	10.8 ± 0.8
	1000	8.8 ± 0.3	12.7 ± 0.6
Ethyl acetate	250	7.8 ± 0.8	9.2 ± 0.3
	500	8.5 ± 0.5	11.5 ± 0.5
	1000	9.3 ± 1.2	14.5 ± 0.5
<i>n</i> -hexane	250	9.3 ± 1.0	7.2 ± 0.3
	500	10.0 ± 0.8	7.7 ± 0.3
	1000	11.7 ± 0.8	8.2 ± 0.3
Salicin	250	7.7 ± 0.3	7.8 ± 0.8
	500	8.2 ± 0.5	8.7 ± 0.6
	1000	8.5 ± 1.2	10.2 ± 0.3
Control (+) <i>Amoxylin</i>	250	16.7 ± 0	27.7 ± 0.6

CONCLUSION

Salicin has been isolated from the ethyl acetate extract of *Salix tetrasperma* Roxb. stem bark and has great potential as a source of the antibacterial compound.

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REFERENCES

- [1] K. R. Kirtikar and B. D. Basu, *Indian Medicinal Plants*. Dehradun: Lalit Mohan Basu, pp. 2362-2363, 2005.
- [2] B. Schmid *et al.*, "Efficacy and tolerability of a standardized willow bark extract in patients with osteoarthritis: randomized placebo-controlled, double blind clinical trial.," *Phytother. Res.*, vol. 15, no. 4, pp. 344–50, Jun. 2001.
- [3] R. Valsaraj, P. Pushpangadan, U. W. Smitt, A. Adersen, and U. Nyman, "Antimicrobial screening of selected medicinal plants from India.," *J. Ethnopharmacol.*, vol. 58, no. 2, pp. 75–83, Oct. 1997.
- [4] A. El-Shazly, A. El-Sayed, and E. Fikrey, "Bioactive secondary metabolites from *Salix tetrasperma* Roxb.," *Zeitschrift fur Naturforschung. C*, vol. 67, no. 7–8, pp. 353–359, 2012.
- [5] E. A. El-Wakil, E.-S. S. Abdel-Hameed, M. M. El-Sayed, and E. E. Abdel-Lateef, "Identification of the chemical composition of the methanolic extract of *Salix tetrasperma* Roxb. using LC-ESI-MS and evaluation its potential as antioxidant agent," *Der Pharma Chem.*, vol. 7, no. 2, pp. 168–177, 2015.

- [6] Y. Fahrimal, A. Rafina, A. Azhar, and D. Nuzul Asmilia, "Blood Profile of Rats (*Rattus norvegicus*) Infected with *Trypanosoma evansi* Treated with Willow Tree Bark Extract (*Salix tetrasperma* Roxb)," *J. Kedokt. Hewan*, vol. 8, no. 2, pp. 164–168, 2014.
- [7] C. R. Raj, GK. Dash, M. Sumanta, and S. Parhi, "Studies on the Hypoglycaemic Activity of The Bark of *Salix tetrasperma* Roxb.," *Int. J. Drug Dev. Res.*, vol. 2, no. 4, 2009.
- [8] M. Sumanta, H. Ramana, and C. Rishi Raj, "Studies on Diuretic and Laxative Activity of the Bark of *Salix Tetrasperma* Roxburgh," *Int. Res. J. Pharm.*, vol. 1, no. 1, pp. 145–149, 2010
- [9] M. S. Islam *et al.*, "Antibacterial, Insecticidal and in Vivocytotoxicity Activities of *Salix Tetrasperma*," *Int. J. Pharm. Sci. Res.*, vol. 2, no. 8, pp. 2103–2108, 2011.
- [10] S. M. Moustafa, B. M. Menshawi, G. M. Wassel, K. Mahmoud, and M. Mounier, "Screening of some Plants in Egypt for their Cytotoxicity against four Human Cancer cell lines," *Int. J. PharmTech Res.*, vol. 6, no. 3, pp. 1074–1084, 2014.
- [11] E. S. Abdel-Hameed, H. A. El-Nahas, and S. A. Abo-Sedera, "Antischistosomal and Antimicrobial Activities of Some Egyptian Plant Species," *Pharm. Biol.*, vol. 46, no. 9, pp. 626–633, Jan. 2008.
- [12] S. A. Deepak, G. Oros, S. G. Sathyanarayana, H. S. Shetty, and S. Sashikanth, "Antisporulant Activity of Watery Extracts of Plants against *Sclerospora graminicola* causing Downy Mildew Disease of Pearl Millet," *Am. J. Agric. Biol. Sci.*, vol. 2, no. 1, pp. 36–42, 2007.
- [13] T. Petkar, A. Siddiqha, and P. T. Kekuda, "Evaluation of antifungal activity of some plants against seed-borne fungi," *Sch. J. Agric. Vet. Sci.*, vol. 4, no. 4, 2017.
- [14] P. T. Kekuda, K.S. Vinayaka and P. S. Kumar, "Antimicrobial activity of *Salix tetrasperma* Roxb. (Salicaceae)," *Int. J. Herb. Med.*, vol. 5, no. 5, pp. 192–195, 2017.
- [15] X. Pan, F. Chen, T. Wu, H. Tang, and Z. Zhao, "The acid, bile tolerance and antimicrobial property of *Lactobacillus acidophilus* NIT," *Food Control*, vol. 20, no. 6, pp. 598–602, Jun. 2009.
- [16] C. Noleto-Dias, J. L. Ward, A. Bellisai, C. Lomax, and M. H. Beale, "Salicin-7-sulfate: A new salicinoid from willow and implications for herbal medicine," *Fitoterapia*, vol. 127, pp. 166–172, Jun. 2018.
- [17] O. Stefanović, I. Radojević, S. Vasić, and L. Čomić, "Antibacterial Activity of Naturally Occurring Compounds from Selected Plants," in *Antimicrobial Agents*, V. Bobbarala, Ed. London: IntechOpen, 2012, pp. 1–25.
- [18] M. O. Soares, A. F. Vinha, F. Coutinho, and P. C. Pires, "Antimicrobial natural products," in *Microbial Pathogens and Strategies for combating them: Science, Technology and Education*, A. Mendez-Vilas, Ed. Elche: Formatex, 2013, pp. 915–916.
- [19] D. Savoia, "Plant-derived antimicrobial compounds: alternatives to antibiotics," *Future Microbiol.*, vol. 7, no. 8, pp. 979–990, Aug. 2012.